

Genome Sequence of *Bartonella birtlesii*, a Bacterium Isolated from Small Rodents of the Genus *Apodemus*

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Bartonella birtlesii is a facultative intracellular bacterium isolated from the blood of small mammals of the genus *Apodemus*. The present study reports the draft genome of *Bartonella birtlesii* strain IBS 135^T (CIP 106691^T).

acteria of the genus Bartonella are facultative intracellular bac-Batteria of the gentle but the state of the gentle but th blood cells as a parasitism strategy (8). Bartonella species belong to the α-2 subgroup of Proteobacteria and are fastidious Gram-negative bacteria highly adapted to their mammalian reservoir hosts, where they live within erythrocytes. To date, more than 30 Bartonella species have been isolated from mammals (including humans and domestic and wild animals), and half of them are considered emerging pathogens (6). Bartonella birtlesii has been isolated for the first time from the blood of small mammals of the genus Apodemus in France and the United Kingdom (1). Although its pathogenicity in humans is unknown, this bacterium has been mainly used as a model to study long-term bacteremia and erythrocyte adhesion and invasion in mice (2, 9). Using signaturetagged mutagenesis of this bacterium, components of the type IV secretion system (T4SS) Trw were identified as the molecular determinant for host-specific erythrocyte infection (9). Therefore, in order to be able to identify any disrupted genes in this signaturetagged mutagenesis library, we sequenced the genome of B. birtlesii.

Genomic DNA from *B. birtlesii* (strain IBS 325^T, CIP 106294^T) was fully sequenced by pyrosequencing using a Titanium genome sequencer (454 Life Sciences, Branford, CT) (5). A library of paired-end fragments was created following the manufacturer's instructions (454 Life Sciences). This library was sequenced using a GS Titanium sequencer (454 Life Sciences) (5).

Reads originated from the strain were assembled into contigs, and a scaffold was created using Newbler 2.53 (454 Life Sciences, Branford, CT). The assembly was verified using CLC Genomics software (CLC Bio, MA). Open reading frame (ORF) prediction was performed on the wild-type strain using PRODIGAL (3). Predicted proteins were compared against the GenBank nonredundant database using BLASTP (http://www.ncbi.nlm.nih.gov) for functional annotation. tRNAs and rRNAs were predicted using ARAGORN (4) and RNAMMER (3), respectively.

B. birtlesii draft genome sequencing generated one scaffold containing 35 contigs with an average coverage of 27-fold. The genome exhibits a total size of 1,833,875 bp and a G+C content of 37%. A total of 1,648 protein-coding ORFs with an average size of 871 bp were identified and cover 78.6% of the genome. Among those genes we found 106 ORFans (6.4% of the total gene content), 42 tRNAs, and two ribosomal operons with the 3 expected

RNA genes (5S, 23S, and 16S). Other notable functional features were two type IV secretion system operons (VirB and Trw). We identified 1,250 orthologous genes between *B. birtlesii* and *Bartonella tribocorum*, the most closely related *Bartonella* species whose genome has been sequenced (7), using a reciprocal best BLAST hit strategy.

Nucleotide sequence accession numbers. The results from this whole-genome shotgun project have been deposited with DDBJ/EMBL/GenBank under the accession number AKIP00000000. The version described in this paper is the first version, AKIP01000000.

ACKNOWLEDGMENT

This research did not benefit from any external funding.

REFERENCES

- Bermond D, et al. 2000. Bartonella birtlesii sp. nov., isolated from small mammals (Apodemus spp.). Int. J. Syst. Evol. Microbiol. 50(Pt 6):1973– 1979.
- Boulouis HJ, et al. 2001. Kinetics of *Bartonella birtlesii* infection in experimentally infected mice and pathogenic effect on reproductive functions. Infect. Immun. 69:5313–5317.
- Lagesen K, et al. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res. 35:3100–3108.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res. 32:11–16.
- Margulies M, et al. 2005. Genome sequencing in microfabricated highdensity picolitre reactors. Nature 437:376–380.
- Rolain JM, et al. 2004. Recommendations for treatment of human infections caused by *Bartonella* species. Antimicrob. Agents Chemother. 48: 1921–1933.
- Saenz HL, et al. 2007. Genomic analysis of Bartonella identifies type IV secretion systems as host adaptability factors. Nat. Genet. 39:1469–1476.
- Saisongkorh W, Rolain JM, Suputtamongkol Y, Raoult D. 2009. Emerging *Bartonella* in humans and animals in Asia and Australia. J. Med. Assoc. Thai. 92:707–731.
- 9. Vayssier-Taussat M, et al. 2010. The Trw type IV secretion system of *Bartonella* mediates host-specific adhesion to erythrocytes. PLoS Pathog. 6:e1000946. doi:10.1371/journal.ppat.1000946.

Received 11 June 2012 Accepted 20 June 2012
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doi:10.1128/JB.01044-12